

# Elevated Plasma Cyclophilin A in Hemodialysis and Peritoneal Dialysis Patients

## A Novel Link to Systemic Inflammation

Kyubok Jin,<sup>1</sup> Nosratola D Vaziri<sup>2</sup>

<sup>1</sup>Division of Nephrology, Department of Internal Medicine, Keimyung University Dongsan Medical Center, Daegu, Korea

<sup>2</sup>Division of Nephrology and Hypertension, University of California, Irvine, California

**Keywords.** cyclophilin A, atherosclerosis, hemodialysis, peritoneal dialysis, chronic kidney disease, inflammation, diabetes mellitus, oxidative stress

**Introduction.** Cyclophilin A has emerged as a novel mediator of oxidative stress and inflammation and a major player in cardiovascular disease, diabetes mellitus, viral infections, and neurodegenerative and thrombotic disorders. Cyclophilin A is released by certain cell types spontaneously or in response to inflammatory mediators, hypoxia, oxidative stress, and hyperglycemia. Many of these conditions are either present or frequently occur in patients with end-stage renal disease and can stimulate release of cyclophilin A, thereby amplifying systemic inflammation. To our knowledge, the effect of end-stage renal disease and dialysis modalities on circulating cyclophilin A has not been previously investigated. This study tested the hypothesis that extracellular cyclophilin A is elevated in patients maintained on hemodialysis and peritoneal dialysis.

**Materials and Methods.** Cyclophilin A, high-sensitivity C-reactive protein, interleukin-6, tumor necrosis factor- $\alpha$ , and lipid levels were measured in the fasting plasma samples from 20 hemodialysis and 20 peritoneal dialysis patients, and 20 age- and sex-matched controls.

**Results.** Plasma cyclophilin A concentration in the patients on hemodialysis ( $105.3 \pm 6.2$  ng/mL) and peritoneal dialysis ( $106.8 \pm 9.0$  ng/mL) were significantly higher than that in the control group ( $29.7 \pm 4.1$  ng/mL). This was associated with significant elevation of high-sensitivity C-reactive protein, interleukin-6, and tumor necrosis factor- $\alpha$ . Plasma cyclophilin A concentration showed direct correlations with high-sensitivity C-reactive protein, interleukin-6, and tumor necrosis factor- $\alpha$ , and an inverse correlation with high-density lipoprotein cholesterol concentration.

**Conclusions.** Plasma cyclophilin A concentration is markedly elevated and positively correlates with the markers of systemic inflammation in hemodialysis and peritoneal dialysis patients.

IJKD 2017;11:44-9  
www.ijkd.org

### INTRODUCTION

Systemic oxidative stress and inflammation are invariably present and play a central part in the pathogenesis of cardiovascular disease, cachexia, anemia, and various other complications in patients with end-stage renal disease (ESRD).<sup>1-6</sup> Several

factors contribute to the ESRD-associated systemic inflammation and oxidative stress including uremic toxins and metabolites, hypervolemia, hypertension, upregulation and activation of tissue angiotensin system, impaired nuclear factor (erythroid-derived 2)-like 2-mediated expression of endogenous

antioxidant and detoxifying molecules,<sup>7-10</sup> disruption of intestinal epithelial barrier structure and function leading to endotoxemia,<sup>11-14</sup> and altered gut microbial flora<sup>15,16,18</sup> among others.

Cyclophilin A has emerged as a novel mediator of oxidative stress and inflammation and a major player in cardiovascular disease, diabetes mellitus, viral infections, neurodegenerative diseases, and thrombotic disorders.<sup>19,20</sup> Cyclophilin A is an abundant intracellular housekeeping protein, which belongs to the cyclophilin family of proteins found in all mammalian cells.<sup>20</sup> Although cyclophilin A is an intracellular protein, a variety of cell types release cyclophilin A spontaneously or in response to inflammatory mediators, hypoxia, oxidative stress, and other stimuli. For instance, macrophages release cyclophilin A following exposure to endotoxin or high glucose concentrations and during their transformation to foam cell<sup>21-23</sup>; cardiomyocytes release cyclophilin A when exposed to hypoxia<sup>24</sup>; and platelets release cyclophilin A upon activation.<sup>25</sup> Release of cyclophilin A to the extracellular space requires its acetylation, which amplifies its pro-inflammatory property.<sup>26</sup> In addition cell death results in the release of cyclophilin A. The extracellular cyclophilin A binds to the extracellular matrix metalloproteinase inducer (CD147), which is widely expressed on cardiovascular cell types.<sup>27</sup> The cyclophilin A-CD147 complex is a powerful chemotactic factor for T cells, monocytes, and neutrophilic and eosinophilic polymorphonuclear leukocytes and promotes activation of nuclear factor kappa-light-chain-enhancer of activated B cells, phosphoinositide 3-kinase, or extracellular signal-regulated kinase in relevant cell types,<sup>28,29</sup> as well as activation of extracellular matrix metalloproteinases.<sup>23,30</sup> There is mounting evidence supporting the role of cyclophilin A-CD147 interaction in endothelial injury and dysfunction,<sup>31,32</sup> vascular remodeling,<sup>33</sup> atherosclerosis,<sup>19,23,25,34</sup> vascular thrombosis,<sup>35</sup> myocardial infarction,<sup>36</sup> and cardiomyopathies.<sup>37,38</sup> In addition, cyclophilin A plays a critical role in replication and release of hepatitis B and C viruses.<sup>39-42</sup>

Many of the conditions known to promote release of extracellular cyclophilin A, ie, oxidative stress, inflammation, hyperglycemia, and ischemia or hypoxia are either present or frequently occur in ESRD patients. As noted above, oxidative stress and inflammation are commonly present

and are frequently accompanied by endotoxemia in patients with ESRD.<sup>43,44</sup> In addition, due to diabetes mellitus, which is the most common cause of ESRD and peritoneal dialysis (PD) procedure with high glucose dialysis solutions, ESRD patients frequently experience episodes of hyperglycemia. Moreover, intradialytic and postdialysis hypotension commonly result in episodes of tissue ischemia and fluid overload, and anemia and dialysis-induced hypoventilation result in hypoxia and impaired oxygen delivery. Together, these conditions can stimulate release and increase plasma concentration of cyclophilin A, which can amplify systemic inflammation and create a vicious circuit. To our knowledge, the effect of ESRD and dialysis modalities on circulating cyclophilin A has not been previously investigated. The present study was undertaken to test the hypothesis that extracellular cyclophilin A is elevated in ESRD patients maintained on hemodialysis and PD.

## MATERIALS AND METHODS

### Study Groups

Twenty hemodialysis patients (10 men and 10 women; mean age, 49.4 ± 11.8 years), and 20 PD patients (10 men and 10 women; mean age, 50.1 ± 8.4 years) were recruited into the study. The underlying causes of kidney disease in the hemodialysis group included diabetes mellitus in 10, hypertension in 8, and chronic glomerulonephritis in 2 patients. The underlying causes of kidney disease in the PD group were diabetes mellitus in 12, hypertension in 5, and chronic glomerulonephritis in 3 patients. Duration of dialysis in the hemodialysis group (24.9 ± 14.3 months) was comparable to that of the PD group (21.6 ± 11.3 months). Twenty age- and sex-matched healthy individuals (10 men and 10 women; mean age, 47.7 ± 8.5 years) served as controls.

Individuals younger than 18 years, those with a history of malignancy or chronic liver disease, and those with a history of infection within the previous 4 weeks were excluded from the study. The study protocol was approved by the Human Subjects Institutional Review Board of the Inje University Haeundae Paik Hospital (129792-2014-045), and all participants signed the informed consent forms.

### Laboratory Measurements

Fasting blood samples were obtained by

venipuncture from all patients and controls. Total cholesterol, low-density lipoprotein cholesterol (LDLC), high-density lipoprotein cholesterol, triglyceride, creatinine, and urea nitrogen were measured by the central laboratory of the Inje University Haeundae Paik Hospital.

Plasma cyclophilin A was measured by an enzyme-linked immunosorbent assay using the kit purchased from Cusabio Biotech (Carlsbad, CA, USA) according to the manufacturer's specifications. High-sensitivity C reactive protein (HSCRP) was measured as a biomarker of systemic inflammation by turbidimetric immunoassay using the kit purchased from Sekisui Chemical (Osaka, Japan) according to the manufacturer's specifications. Interleukin-6 (IL-6) and tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) were measured by an enzyme-linked immunosorbent assay using the kit purchased from R&D Systems (Minneapolis, MN, USA).

### Statistical Analysis

The analysis of variance and a regression model were used in statistical analysis of the data. Continuous variables were expressed as mean  $\pm$  standard error of the mean. *P* values less than .05 were considered significant.

## RESULTS

### Characteristics of Study Groups

Data are shown in Table 1. Arterial blood pressure in hemodialysis and PD groups were comparable

and significantly higher than that found in the control group. Blood hemoglobin concentrations in the hemodialysis and PD groups were comparable and significantly lower than that found in the control group. Serum albumin concentration in the PD group was significantly lower than in the hemodialysis group and significantly lower in both than in the controls. Serum calcium was significantly lower whereas serum phosphorus and calcium-phosphorus product were significantly higher in the hemodialysis and PD groups compared with the corresponding values found in the control group. Serum urea nitrogen and creatinine levels were markedly elevated and serum uric acid was modestly increased in the hemodialysis and PD groups as compared to the corresponding values found in the control group. Serum total cholesterol, LDLC, and triglyceride concentrations were significantly higher in the PD group than in the hemodialysis and control groups. Serum high-density lipoprotein cholesterol was significantly lower in both hemodialysis and PD groups than in the control group.

### Plasma Cyclophilin A and Inflammatory Markers

The laboratory results are shown in Table 2. Plasma cyclophilin A concentration in the hemodialysis and peritoneal dialysis groups were significantly higher than that in the control group (*P* < .001). This was associated with significant elevation

**Table 1.** Laboratory and Clinical Measures in Hemodialysis, Peritoneal Dialysis, and Control Groups

Parameter	Control Group (n = 20)	Hemodialysis Group (n = 20)	Peritoneal Dialysis Group (n = 20)
Systolic blood pressure, mmHg	118.9 $\pm$ 7.2	140.7 $\pm$ 17.8*	138.5 $\pm$ 26.4*
Diastolic blood pressure, mmHg	76.8 $\pm$ 8.1	85.5 $\pm$ 15.1*	84.0 $\pm$ 13.1*
Hemoglobin, g/dL	14.2 $\pm$ 1.6	10.1 $\pm$ 0.8*	10.2 $\pm$ 1.1
Serum albumin, g/dL	4.5 $\pm$ 0.2	3.6 $\pm$ 0.3*	3.2 $\pm$ 0.5*§
Calcium, mmol/L	9.5 $\pm$ 0.4	8.3 $\pm$ 0.5†	8.2 $\pm$ 0.7*
Phosphorus, mmol/L	35 $\pm$ 0.8	5.9 $\pm$ 2.2*	5.4 $\pm$ 1.9*
Calcium-phosphorus product, mg <sup>2</sup> /dL <sup>2</sup>	33.2 $\pm$ 8.6	48.5 $\pm$ 17.3*	44.2 $\pm$ 15.3*
Serum urea nitrogen, mg/dL	11.5 $\pm$ 2.8	76.9 $\pm$ 29.0*	57.2 $\pm$ 16.2*
Serum creatinine, mg/dL	0.9 $\pm$ 0.2	10.6 $\pm$ 2.6*	10.6 $\pm$ 4.6*
Total cholesterol, mg/dL	178.4 $\pm$ 34.1	153.1 $\pm$ 31.3†	212.3 $\pm$ 30.8*§
Low-density lipoprotein cholesterol, mg/dL	85.1 $\pm$ 27.4	88.3 $\pm$ 24.5†	135.8 $\pm$ 19.9*§
High-density lipoprotein cholesterol, mg/dL	54.3 $\pm$ 111.1	40.1 $\pm$ 9.9†	36.6 $\pm$ 8.9*
Triglyceride, mg/dL	93.5 $\pm$ 45.6	113.5 $\pm$ 64.5	194.0 $\pm$ 97.6*‡
Uric acid, mg/dL	4.2 $\pm$ 1.2	7.4 $\pm$ 1.5*	7.3 $\pm$ 1.5*

\**P* < 0.01 versus the control group

†*P* < 0.05 versus the control group

‡*P* < 0.01 versus the hemodialysis group

§*P* < 0.05 versus the hemodialysis group

**Table 2.** Plasma Cyclophilin A, High-sensitivity C-Reactive Protein, Interleukin-6, and Tumor Necrosis Factor- $\alpha$  Concentrations in Hemodialysis, Peritoneal Dialysis and Control Groups

Parameter	Control Group (n = 20)	Hemodialysis Group (n = 20)	Peritoneal Dialysis Group (n = 20)
Cyclophilin A, ng/mL	29.7 $\pm$ 4.1	105.3 $\pm$ 6.2*	106.8 $\pm$ 9.0*
High-sensitivity C-reactive protein, mg/dL	0.03 $\pm$ 0.01	0.29 $\pm$ 0.29*	0.24 $\pm$ 0.24*
Interleukin-6, pg/mL	1.3 $\pm$ 0.2	5.2 $\pm$ 0.4*	5.2 $\pm$ 0.5*
Tumor necrosis factor- $\alpha$ , pg/mL	1.4 $\pm$ 0.1	4.6 $\pm$ 0.2*	4.8 $\pm$ 0.3*

\* $P < 0.01$  versus the control group

of plasma HSCRP, IL-6, and TNF- $\alpha$ . There was a significant direct correlation between plasma cyclophilin A and HSCRP ( $r = 0.436$ ,  $P = .01$ ), IL-6 ( $r = 0.526$ ,  $P = .001$ ), TNF- $\alpha$  ( $r = 0.651$ ,  $P = .001$ ), systolic blood pressure ( $r = 0.386$ ,  $P = .01$ ), and diastolic blood pressure ( $r = 0.360$ ,  $P = 0.05$ ), and an inverse relationship between cyclophilin A and plasma high-density lipoprotein cholesterol concentration ( $r = -0.634$ ,  $P = 0.01$ ) in the study population.

## DISCUSSION

The present study revealed a marked elevation of plasma cyclophilin A levels in stable patients with ESRD maintained on hemodialysis and PD modalities. Plasma cyclophilin A concentration showed a direct correlation with HSCRP, IL-6, and TNF- $\alpha$ , the well-known markers of systemic inflammation. This finding is consistent with participation of cyclophilin A and systemic inflammation in a vicious circuit wherein inflammation promotes release of cyclophilin A and cyclophilin A amplifies the inciting inflammation.

As noted in the introduction, emerging data have demonstrated the role of cyclophilin A in the pathogenesis of atherosclerosis, arteriosclerosis, and cardiovascular disease,<sup>19,37,47</sup> which are the main causes of morbidity and mortality in chronic kidney disease (CKD) population. The role of cyclophilin A in the pathogenesis of arteriosclerosis and arterial remodeling has been demonstrated in several studies conducted in experimental animals. Using carotid artery ligation model, Satoh and coworkers found significant protection against intimal and medial hyperplasia in genetically cyclophilin A deficient compared to the wild type mice.<sup>33</sup> This was associated with diminished vascular smooth muscle cell proliferation and reduced monocyte infiltration, which are critical steps in arteriosclerosis and atherosclerosis. Likewise, cyclophilin A-deficient

mice have been shown to be completely protected against angiotensin-2 induced abdominal aortic aneurysm.<sup>46</sup> This phenomenon is attributed to attenuation of tissue inflammation and oxidative stress and suppression of metalloproteinases, which are essential for aneurysm formation.

Compared with the hemodialysis group, the PD patients exhibited significant increase in serum total and LDLC and triglyceride concentrations. This phenomenon is caused by significant daily losses of protein in the effluent peritoneal dialysis fluid which simulate nephrotic syndrome in these functionally anephric patients.<sup>47,48</sup>

Cyclophilin A has a peptidylprolyl-cis/trans isomerase enzymatic activity which regulates intracellular folding and trafficking of proteins. Cyclophilin A was first identified in 1984 to be the intracellular protein which binds the immunosuppressive drug, cyclosporine A. Cyclophilin A was shown to bind cyclosporine A and form a complex that can suppress immune response by blocking calcineurin-dependent activation of nuclear factor activating T lymphocytes.<sup>49</sup> Due to its potent immunosuppressive property and other side effects cyclosporine A is not suitable for the treatment of systemic inflammation in patients with CKD, type-2 diabetes mellitus, atherosclerosis, and other chronic illnesses. Presently, drugs that can exclusively block extracellular cyclophilin A and lack immunosuppressive properties are not available. Development of such agents will undoubtedly prove highly effective in the management of chronic inflammatory diseases including CKD.

Due to occasional blood transfusion and surgical procedures, patients with advanced CKD are at a high risk of exposure to viral hepatitis. Given the central role of cyclophilin A in proliferation and release of hepatitis B and C viruses,<sup>39-42</sup> elevation of cyclophilin A level in hemodialysis and peritoneal

dialysis patients shown here may impact the course and outcomes of these infections.

Given the role of extracellular cyclophilin A as a biomarker and potent mediator of oxidative stress, inflammation, and cardiovascular disease, which are the common features of CKD, cyclophilin A is an attractive diagnostic and therapeutic target in CKD population.

The authors wish to acknowledge the limitations of the study including the relatively small size of the enrolled populations and cross-sectional nature of the study. Longitudinal studies enrolling larger number of participants are needed to confirm the results of the present study and to determine the impact of elevated cyclophilin A on morbidity and mortality in CKD populations.

## CONCLUSIONS

Plasma cyclophilin A concentration is markedly elevated and positively correlates with the markers of systemic inflammation and cardiovascular disease in hemodialysis and peritoneal dialysis patients.

## ACKNOWLEDGEMENTS

This research was supported by the Keimyung University Research Grant of 2016.

## CONFLICT OF INTEREST

None declared.

## REFERENCES

1. Stenvinkel P. Inflammation in end-stage renal disease--a fire that burns within. *Contrib Nephrol.* 2005;149:185-99.
2. Himmelfarb J, Stenvinkel P, Ikizler TA, Hakim RM. The elephant in uremia: oxidant stress as a unifying concept of cardiovascular disease in uremia. *Kidney Int.* 2002;62:1524-38.
3. Vaziri ND. Oxidative stress in uremia: nature, mechanisms, and potential consequences. *Semin Nephrol.* 2004;24:469-73.
4. Cachofeiro V, Goicochea M, de Vinuesa SG, Oubina P, Lahera V, Luno J. Oxidative stress and inflammation, a link between chronic kidney disease and cardiovascular disease. *Kidney Int Suppl.* 2008:S4-9.
5. Yoon JW, Pahl MV, Vaziri ND. Spontaneous leukocyte activation and oxygen-free radical generation in end-stage renal disease. *Kidney Int.* 2007;71:167-72.
6. Kaysen GA. Biochemistry and biomarkers of inflamed patients: why look, what to assess. *Clin J Am Soc Nephrol.* 2009;4 Suppl 1:S56-63.
7. Kim HJ, Vaziri ND. Contribution of impaired Nrf2-Keap1 pathway to oxidative stress and inflammation in chronic renal failure. *Am J Physiol Renal Physiol.* 2010;298:F662-71.
8. Kim HJ, Sato T, Rodriguez-Iturbe B, Vaziri ND. Role of intrarenal angiotensin system activation, oxidative stress, inflammation, and impaired nuclear factor-erythroid-2-related factor 2 activity in the progression of focal glomerulosclerosis. *J Pharmacol Exp Ther.* 2011;337:583-90.
9. Aminzadeh MA, Nicholas SB, Norris KC, Vaziri ND. Role of impaired Nrf2 activation in the pathogenesis of oxidative stress and inflammation in chronic tubulo-interstitial nephropathy. *Nephrol Dial Transplant.* 2013;28:2038-45.
10. Ruiz S, Pergola PE, Zager RA, Vaziri ND. Targeting the transcription factor Nrf2 to ameliorate oxidative stress and inflammation in chronic kidney disease. *Kidney Int.* 2013;83:1029-41.
11. Ritz E. Intestinal-renal syndrome: mirage or reality? *Blood Purif.* 2011;31:70-6.
12. Vaziri ND, Yuan J, Rahimi A, Ni Z, Said H, Subramanian VS. Disintegration of colonic epithelial tight junction in uremia: a likely cause of CKD-associated inflammation. *Nephrol Dial Transplant.* 2012;27:2686-93.
13. Vaziri ND, Yuan J, Nazertehrani S, Ni Z, Liu S. Chronic kidney disease causes disruption of gastric and small intestinal epithelial tight junction. *Am J Nephrol.* 2013;38:99-103.
14. Vaziri ND, Goshtasbi N, Yuan J, et al. Uremic plasma impairs barrier function and depletes the tight junction protein constituents of intestinal epithelium. *Am J Nephrol.* 2012;36:438-43.
15. Vaziri ND, Wong J, Pahl M, et al. Chronic kidney disease alters intestinal microbial flora. *Kidney Int.* 2013;83:308-15.
16. Wong J, Piceno YM, Desantis TZ, Pahl M, Andersen GL, Vaziri ND. Expansion of urease- and uricase-containing, indole- and p-cresol-forming and contraction of short-chain fatty acid-producing intestinal microbiota in ESRD. *Am J Nephrol.* 2014;39:230-7.
17. Vaziri ND, Zhao YY, Pahl MV. Altered intestinal microbial flora and impaired epithelial barrier structure and function in CKD: the nature, mechanisms, consequences and potential treatment. *Nephrol Dial Transplant.* 2016;31:737-46.
18. Nigro P, Pompilio G, Capogrossi MC. Cyclophilin A: a key player for human disease. *Cell Death Dis.* 2013;4:e888.
19. Satoh K. Cyclophilin A in Cardiovascular Homeostasis and Diseases. *Tohoku J Exp Med.* 2015;235:1-15.
20. Marks WH, Harding MW, Handschumacher R, Marks C, Lorber MI. The immunochemical distribution of cyclophilin in normal mammalian tissues. *Transplantation.* 1991;52:340-5.
21. Ramachandran S, Venugopal A, Sathisha K, et al. Proteomic profiling of high glucose primed monocytes identifies cyclophilin A as a potential secretory marker of inflammation in type 2 diabetes. *Proteomics.* 2012;12:2808-21.
22. Sherry B, Yarlett N, Strupp A, Cerami A. Identification of cyclophilin as a proinflammatory secretory product of lipopolysaccharide-activated macrophages. *Proc Natl Acad Sci U S A.* 1992;89:3511-5.
23. Seizer P, Schonberger T, Schott M, et al. EMMPRIN and its ligand cyclophilin A regulate MT1-MMP, MMP-9

- and M-CSF during foam cell formation. *Atherosclerosis*. 2010;209:51-7.
24. Seko Y, Fujimura T, Taka H, Mineki R, Murayama K, Nagai R. Hypoxia followed by reoxygenation induces secretion of cyclophilin A from cultured rat cardiac myocytes. *Biochem Biophys Res Commun*. 2004;317:162-8.
  25. Coppinger JA, Cagney G, Toomey S, et al. Characterization of the proteins released from activated platelets leads to localization of novel platelet proteins in human atherosclerotic lesions. *Blood*. 2004;103:2096-104.
  26. Soe NN, Sowden M, Baskaran P, et al. Acetylation of cyclophilin A is required for its secretion and vascular cell activation. *Cardiovasc Res*. 2014;101:444-53.
  27. Damsker JM, Bukrinsky MI, Constant SL. Preferential chemotaxis of activated human CD4+ T cells by extracellular cyclophilin A. *J Leukoc Biol*. 2007;82:613-8.
  28. Schmidt R, Bultmann A, Fischel S, et al. Extracellular matrix metalloproteinase inducer (CD147) is a novel receptor on platelets, activates platelets, and augments nuclear factor kappaB-dependent inflammation in monocytes. *Circ Res*. 2008;102:302-9.
  29. Bahmed K, Henry C, Holliday M, et al. Extracellular cyclophilin-A stimulates ERK1/2 phosphorylation in a cell-dependent manner but broadly stimulates nuclear factor kappa B. *Cancer Cell Int*. 2012;12:19.
  30. Yurchenko V, Constant S, Bukrinsky M. Dealing with the family: CD147 interactions with cyclophilins. *Immunology*. 2006;117:301-9.
  31. Nigro P, Satoh K, O'Dell MR, et al. Cyclophilin A is an inflammatory mediator that promotes atherosclerosis in apolipoprotein E-deficient mice. *J Exp Med*. 2011;208:53-66.
  32. Jin ZG, Lungu AO, Xie L, Wang M, Wong C, Berk BC. Cyclophilin A is a proinflammatory cytokine that activates endothelial cells. *Arterioscler Thromb Vasc Biol*. 2004;24:1186-91.
  33. Satoh K, Matoba T, Suzuki J, et al. Cyclophilin A mediates vascular remodeling by promoting inflammation and vascular smooth muscle cell proliferation. *Circulation*. 2008;117:3088-98.
  34. Major TC, Liang L, Lu X, Rosebury W, Bocan TM. Extracellular matrix metalloproteinase inducer (EMMPRIN) is induced upon monocyte differentiation and is expressed in human atheroma. *Arterioscler Thromb Vasc Biol*. 2002;22:1200-7.
  35. Pennings GJ, Yong AS, Kritharides L. Expression of EMMPRIN (CD147) on circulating platelets in vivo. *J Thromb Haemost*. 2010;8:472-81.
  36. Seizer P, Ochmann C, Schonberger T, et al. Disrupting the EMMPRIN (CD147)-cyclophilin A interaction reduces infarct size and preserves systolic function after myocardial ischemia and reperfusion. *Arterioscler Thromb Vasc Biol*. 2011;31:1377-86.
  37. Seizer P, Geisler T, Bigalke B, et al. EMMPRIN and its ligand cyclophilin A as novel diagnostic markers in inflammatory cardiomyopathy. *Int J Cardiol*. 2013;163:299-304.
  38. Zuern CS, Muller KA, Seizer P, et al. Cyclophilin A predicts clinical outcome in patients with congestive heart failure undergoing endomyocardial biopsy. *Eur J Heart Fail*. 2013;15:176-84.
  39. Liu Z, Yang F, Robotham JM, Tang H. Critical role of cyclophilin A and its prolyl-peptidyl isomerase activity in the structure and function of the hepatitis C virus replication complex. *J Virol*. 2009;83:6554-65.
  40. Coelmont L, Hanoulle X, Chatterji U, et al. DEB025 (Alisporivir) inhibits hepatitis C virus replication by preventing a cyclophilin A induced cis-trans isomerisation in domain II of NS5A. *PLoS One*. 2010;5:e13687.
  41. Chatterji U, Bobardt M, Selvarajah S, et al. The isomerase active site of cyclophilin A is critical for hepatitis C virus replication. *J Biol Chem*. 2009;284:16998-7005.
  42. Tian X, Zhao C, Zhu H, et al. Hepatitis B virus (HBV) surface antigen interacts with and promotes cyclophilin A secretion: possible link to pathogenesis of HBV infection. *J Virol*. 2010;84:3373-81.
  43. Szeto CC, Kwan BC, Chow KM, et al. Endotoxemia is related to systemic inflammation and atherosclerosis in peritoneal dialysis patients. *Clin J Am Soc Nephrol*. 2008;3:431-6.
  44. Feroze U, Kalantar-Zadeh K, Sterling KA, et al. Examining associations of circulating endotoxin with nutritional status, inflammation, and mortality in hemodialysis patients. *J Ren Nutr*. 2012;22:317-26.
  45. Seizer P, Gawaz M, May AE. Cyclophilin A and EMMPRIN (CD147) in cardiovascular diseases. *Cardiovasc Res*. 2014;102:17-23.
  46. Satoh K, Nigro P, Matoba T, et al. Cyclophilin A enhances vascular oxidative stress and the development of angiotensin II-induced aortic aneurysms. *Nat Med*. 2009;15:649-56.
  47. Vaziri ND. Causes of dysregulation of lipid metabolism in chronic renal failure. *Semin Dial*. 2009;22:644-51.
  48. Jin K, Park BS, Kim YW, Vaziri ND. Plasma PCSK9 in nephrotic syndrome and in peritoneal dialysis: a cross-sectional study. *Am J Kidney Dis*. 2014;63:584-9.
  49. Handschumacher RE, Harding MW, Rice J, Drugge RJ, Speicher DW. Cyclophilin: a specific cytosolic binding protein for cyclosporin A. *Science*. 1984;226:544-7.

Correspondence to:

Kyubok Jin, MD, PhD  
 Division of Nephrology, Department of Internal Medicine,  
 Keimyung University Dongsan Medical Center  
 56 Dalseong-ro, Jung-gu, Daegu, 700-712, Korea  
 Tel: +82 53 250 7913  
 E-mail: mdjin922@gmail.com

Received April 2016  
 Revised August 2016  
 Accepted September 2016