Effects of Omega-3 Fatty Acid Supplementation on Inflammatory Cytokines and Advanced Glycation End Products in Patients With Diabetic Nephropathy A Randomized Controlled Trial

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Introduction. This study was performed to evaluate the effects of omega-3 fatty acid supplementation on inflammatory cytokines and advanced glycation end products (AGEs) in patients with diabetic nephropathy (DN).

Materials and Methods. This randomized double-blind placebocontrolled trial was done on 60 patients with DN who were randomly divided into 2 groups to receive either 1000 mg/d of omega-3 fatty acid from flaxseed oil (n = 30) or placebo (n = 30) for 12 weeks. The primary outcome variables were tumor necrosis factor- α , receptor tumor necrosis factor- α and growth differentiation factor 15. Fasting blood samples were taken at the onset and the end of the study to quantify the related markers.

Results. Compared with the placebo, omega-3 fatty acid supplementation resulted in a significant decrease in serum AGEs (-2.3 ± 2.8 AU versus 0.2 ± 2.5 AU, *P* = .001). Despite a significant reduction in serum level of receptor for AGEs (-0.1 ± 0.3 AU, *P* = .02) in the omega-3 fatty acid group, no significant difference was found between the two groups in terms of their effects on the receptor for AGEs. Supplementation with omega-3 fatty acid had no significant effect on the inflammatory cytokines as compared with the placebo.

Conclusions. Our study demonstrated that omega-3 fatty acid supplementation among DN patients had favorable effects on AGEs and the receptor for AGEs.

IJKD 2016;10:197-204 www.ijkd.org

INTRODUCTION

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inflammatory markers,

products

advanced glycation end

Keywords. omega-3 fatty

acids, diabetic nephropathy,

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Diabetic nephropathy (DN) is one of the most devastating complications of diabetes mellitus and accounts for a significant increase in morbidity and mortality.¹ Previous studies have demonstrated that inflammatory cytokines, including tumor necrosis factor- α (TNF- α) is directly involved in the pathogenesis of DN by tissue damage and albuminuria related to a enhanced stimuli for overexpression of TNF- α .² In addition, the accumulation of advanced glycation end products (AGEs) and increased inflammatory cytokines play an important role in diabetes complications.^{3,4} Extracellular AGEs, through their interaction with receptor for AGEs (RAGEs), and intracellular AGEs can result in increased oxidative stress and generation of reactive oxygen species and activation of protein kinase C.⁵

Despite recent advancements in improving inflammatory cytokines and AGEs of DN patients, no single medication has thus far been able to correct inflammatory state and AGEs. Several published studies have recently exhibited the beneficial effects of omega-3 fatty acids supplementation in DN patients, with most focusing on metabolic profiles.^{6,7} However, some studies have evaluated the effects of omega-3 fatty acids supplementation on inflammatory cytokines in patients without DN, the results of which are conflicting. It is well established that omega-3 fatty acids supplementation can improve inflammatory cytokines and RAGEs expression.^{8,9} However, some researchers did not observe the beneficial effects of omega-3 fatty acids administration on inflammatory markers.^{10,11} These discrepancies in various studies may be explained by the different population studies, the origin of omega-3 fatty acids, dosage of used omega-3 fatty acids, and duration of treatment.

Omega-3 fatty acids intake may reduce inflammatory cytokines and AGEs through nuclear factor kappa-light-chain-enhancer of activated B cells (NF-kB) activation by increasing peroxisome proliferator-activated receptor- γ (PPAR- γ) gene expression in a chain length-dependent manner.¹²⁻¹⁴ We hypothesized that omega-3 fatty acids supplementation may have a beneficial effect on inflammatory cytokines, AGEs, and RAGEs in patients with DN. This clinical trial was to assay the effects of omega-3 fatty acid supplementation on inflammatory cytokines, AGEs and RAGEs among DN patients.

MATERIALS AND METHODS Participants

In a randomized double-blind placebo-controlled trial, we selected 60 patients with DN referred to Akhavan Clinic in Kashan, Iran, from April 2015 to July 2015. We defined DN as diabetic renal disease with a proteinuria level greater than 0.3 g/24 h, with or without elevation of serum creatinine levels.¹⁵ To determine the sample size, we used the standard formula suggested for clinical trials considering a type 1 error of 0.05 and a type 2 error of 0.20 (power = 80%). Based on a previous randomized controlled trial,⁸ we used 0.20 ng/ mL as the standard deviation and 0.16 ng/mL as the change in the mean value of TNF- α as the key variable. Based on these, we planned to reach 25 patients in each group. Assuming 5 dropouts in each group, the final sample size was determined to be 30 participants per group.

Inclusion and Exclusion Criteria

Eligible participants were patients with DN aged 45 to 85 years old. The exclusion criteria were the intake of omega-3 fatty acid supplements within the past 3 months, uncontrolled diabetes mellitus (hemoglobin A1c [HbA1c], > 7.5%; fasting plasma glucose, > 126 mg/dL; and blood glucose 2-hour postprandial, > 180 mg/dL), current use of warfarin, malignancy, liver cirrhosis, and smoking.

Ethics Statements

This study was conducted in accordance with the Declaration of Helsinki. Participants were informed about the nature and purpose of the study and signed an informed consent form. The study protocol was approved by the ethics committee of Kashan University of Medical Sciences and was registered in the Iranian Registry of Clinical Trials (IRCT201504095623N41).

Study Design

The participants were randomly divided into 2 groups to receive either 1000 mg/d of omega-3 fatty acid from flaxseed oil or placebo for 12 weeks. The appearance of the placebo was indistinguishable in color, shape, size, packaging, smell, and taste from the omega-3 fatty acid capsule. Omega-3 fatty acid supplements and placebos were produced by Barij Pharmaceutical Company (Kashan, Iran). Randomization assignment was performed by the use of computer-generated random numbers. Randomization and allocation were concealed from the researchers and participants until the main analyses were completed. The randomized allocation sequence, enrolling, and allocating participants to the interventions were conducted by a trained nutritionist at the clinic. At the onset of the study, the participants were requested not to change their routine physical activity or usual dietary intakes throughout the study and not to consume any supplements other than the one provided to them by the investigators as well as not to take any medications that might affect the study results during the 12-week intervention. All of the patients completed 3-day food records and 3 physical activity records at the study baseline, during the study (weeks 3, 6, and 9), and at the end of the study. Physical activity was defined as metabolic equivalents in hours per day. To quantify the metabolic equivalents for each patient, we multiplied the times (in hour per day) reported for each physical activity by its related metabolic equivalents coefficient by standard tables.¹⁶ Daily macro- and micro-nutrient intakes were analyzed by nutritionist IV software (First Databank, San Bruno, CA).

Treatment Adherence

Every 4 weeks, the patients were given enough supplements to last 3 days after their next scheduled visit and were instructed to return all unused supplements at each visit. The remaining supplements were counted and subtracted from the number provided to determine the number taken. To increase the compliance, all of the patients were receiving short messages on their cell phones to take the supplements every day.

Assessment of Anthropometric Measures

Weight and height of the patients were recorded in an overnight fasting status using a standard scale (Seca, Hamburg, Germany) at study baseline and after a 12-week treatment period. All anthropometric measures were conducted by a trained nutritionist who was blinded to the randomization assignments.

Outcomes

The primary outcome variables were inflammatory cytokines, and the secondary outcome variables were AGEs and RAGEs concentrations. Before the onset and after 12 weeks of intervention, 5-mL blood samples were taken in a fasting status from each participant at the Kashan University of Medical Sciences Reference Laboratory. To separate the serum, blood samples were immediately centrifuged (Hettich D-78532, Tuttlingen, Germany) at 3500 rpm for 10 minutes. Then, we stored the samples at -70°C before the final analysis. Serum TNF-α and TNF-α receptor levels were quantified using an enzyme-linked immunosorbent assay kit (Bioassy Technology Laboratory, Shanghai, China) with inter- and intra-assay coefficient variances of 6.5% and 8.5% for TNF- α and 6.7% and 8.9% for TNF- α receptor, respectively. Serum growth differentiation factor 15 concentrations were determined by the use of an enzyme-linked immunosorbent assay kit (Bioassy Technology Laboratory, Shanghai, China) with inter- and intra-assay coefficient variances of 7.1% and 9.3%, respectively. Serum AGEs were assessed by the fluorometeric method with interand intra-assay coefficient variances of 3.9% and 4.8%. Serum RAGEs concentrations were quantified using an enzyme-linked immunosorbent assay kit (Bioassy Technology Laboratory, Shanghai, China) with inter- and intra-assay coefficient variances of 7.3% and 9.4%, respectively. The HbA1c levels in the whole blood were determined by a Glycomat kit (BiocodeHycel, Massy, France) using the method of exchange chromatography. However, effects of omega-3 fatty acid supplementation on HbA1c levels were not evaluated. Data on HbA1c at study baseline were taken from the records of the patients available at the clinic. Enzymatic kits (Pars Azmun, Tehran, Iran) were used to determine fasting plasma glucose and serum creatinine concentrations (Jaffe method) with intra- and inter-assay coefficient variances less than 5%.

Statistical Analyses

We conducted the Kolmogrov-Smirnov test to assess the normality of distribution of variables. The analyses were performed based on the intention-to-treat principle. To detect differences in anthropometric measures as well as in daily dietary intakes between the two groups, we used independent samples Student t test. Data on AGEs, RAGEs, and inflammatory cytokines are presented as mean ± standard deviation. To determine the effects of omega-3 fatty acid administration on AGEs, RAGEs, and inflammatory cytokines, we used the 1-way repeated measures analysis of variance. In this analysis, the treatment (omega-3 fatty acid versus placebo group) was regarded as the between-subject factor and time, with 2 timepoints (the beginning of the study and after 12-week intervention), was considered as the within-subject factor. To examine if age, baseline body mass index and baseline values of the biochemical parameters influenced our findings, we applied the analysis of covariance controlling for these variables. P values less than .05 were considered significant. All statistical analyses were done by the use of the Statistical Package for Social Science, version 18 (SPSS Inc, Chicago, Illinois, USA).

RESULTS

At the screening visit, 205 diabetic patients were screened, of whom 70 had DN. Ten patients did not meet the criteria and 60 patients were enrolled and assigned to the two groups. Each group included 30

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patients, but 2 from each group were lost to follow up (Figure). Finally, 56 participants completed the study (28 in the omega-3 and 28 in the placebo group). In accordance with the intention-to-treat principle, all of the 60 patients were included in the final analysis.

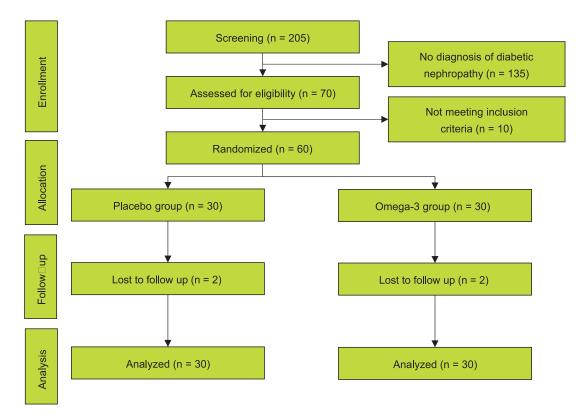
The two groups were comparable in terms of age, height, weight at study baseline and end of the study, weight change, duration of diabetes mellitus, and administration of antidiabetic and antilipidemic medications, angiontensin-converting enzyme inhibitors, and aldosterone receptor blockers (Table 1). Based on the 3-day dietary records obtained at the study baseline and on weeks 3, 6, 9, and 12, no significant changes were observed in the daily dietary macro- and micronutrient intakes between the two groups (Table 2).

After 12 weeks of treatment, omega-3 fatty acid supplements compared with the placebo resulted in a significant decrease in serum AGEs levels (-2.3 ± 2.8 AU versus 0.2 ± 2.5 AU, P = .001; Table 3). Despite a significant reduction of serum RAGEs levels (-0.1 ± 0.3 AU, P = .02) in the omega-3 group, no significant difference was found between the two groups in terms of their effects on RAGEs. Supplementation with omega-3 fatty acid had no significant effect on the inflammatory cytokines compared with the placebo. Intake of omega-3 fatty acid did not significantly affect fasting blood glucose levels compared with the placebo (-3.1 ± 50.5 mg/ dL versus $4.8 \pm 46.2 \text{ mg/dL}$, P = .52). Adjustments for baseline values of biochemical parameters did not affect the findings, except for serum RAGEs (P = .002; Table 4). Further adjustments for age and baseline body mass index did not influence the findings except for serum RAGEs (P = .002).

DISCUSSION

In the current study, we evaluated the effects of omega-3 fatty acid supplementation on AGEs and inflammatory cytokines in patients with DN. However we have previously reported the beneficial effects of omega-3 fatty acid supplementation on markers of insulin resistance and triglycerides levels in patients with DN,⁶ to our knowledge, this randomized clinical trial is the first evaluating the effects of omega-3 fatty acid supplementation on AGEs and inflammatory cytokines these patients.

Patients with DN are susceptible to increased inflammatory cytokines and oxidative stress.^{2,17}



Summary of patient flow diagram.

 Table 1. Characteristics of Study Participants

Characteristic	Placebo Group (n = 30)	Omega-3 Group (n = 30)	Р
Sex, %			
Male	10 (33.3)	11 (36.7)	
Female	20 (66.7)	19 (63.3)	.78
Duration of diabetes mellitus, y	15.2 ± 3.2	15.8 ± 3.0	.48
Age, y	63.1 ± 8.7	63.5 ± 10.1	.86
Height, cm	162.5 ± 7.4	165.0 ± 7.4	.20
Weight at study baseline, kg	79.5 ± 9.2	80.2 ± 13.5	.35
Weight change, kg	0.1 ± 0.5	0.1 ± 0.4	.89
Antidiabetic medication, %			
Metformin	3 (10.0)	3 (10.0)	
Metformin and gliclazide	14 (46.7)	13 (43.3)	
Metformin, gliclazide, and repaglinide	13 (43.3)	14 (46.7)	.96
Antilipidemic therapy, %	25 (82.5)	25 (82.5)	> .99
Antilipidemic medication, %			
Statins	19 (76.0)	17 (68.0)	
Fibrates	2 (8.0)	3 (12.0)	
Statins and fibrates	4 (16.0)	5 (20.0)	.81
Angiotensin-converting enzyme inhibitors, %	26 (86.7)	26 (86.7)	> .99
Angiotensin receptor blockers, %	4 (13.3)	5 (16.7)	.78
Hemoglobin A1c,%	6.8 ± 0.4	6.9 ± 0.4	.77
Urinary albumin excretion, mg/d	496.0 ± 68.0	512.3 ± 63.9	.34
Serum creatinine, mg/d	1.3 ± 0.5	1.4 ± 0.6	.79

Table 2. Dietary Intakes of Study Participants Throughout the Study

Dietary Intake	Placebo Group (n = 30)	Omega-3 Group (n = 30)	Р
Energy, kcal/d	2175 ± 226	2256 ± 225	.17
Fat, g/d	74.4 ± 11.3	77.3 ± 14.9	.40
Saturated fatty acids, g/d	23.5 ± 4.6	24.5 ± 4.9	.44
Polyssaturated fatty acids, g/d	22.0 ± 5.2	23.5 ± 6.2	.34
Monossaturated fatty acids, g/d	20.5 ± 5.1	21.1 ± 5.8	.65
Cholesterol, mg/d	192.9 ± 88.7	189.3 ± 94.3	.88
Omega-3 fatty acids, g/d	0.2 ± 0.1	0.2 ± 0.1	.23
Omega-6 fatty acids, g/d	20.6 ± 4.3	21.7 ± 5.1	.38
Selenium, µg/d	53.8 ± 5.7	53.6 ± 5.0	.86
Magnesium, mg/d	263.2 ± 44.5	279.7 ± 52.1	.19

Although limited data are available evaluating effects of omega-3 fatty acid supplementation in human models. Wang and colleagues¹⁸ demonstrated that docosahexaenoic acid inhibited AGEs-induced microglia activation in in vitro microglia culture system. In addition, 2 individual studies have indicated that the transcription of downstream genes such as RAGE and TNF- α were inhibited with docosahexaenoic acid treatment.^{19,20} These findings indicated that docosahexaenoic acid intake may efficiently inhibit NF-kB activity. These results are consistent with other studies in

skeletal muscle and cardiomyocytes,¹⁹ showing eicosapentaenoic acid and docosahexaenoic acid inhibited NF-kB activation by preventing I kappa B alpha phosphorylation and further inhibited NF-kB activation by reducing degradation of the inhibitory I kappa B alpha protein. Furthermore, it was noteworthy that omega-3 fatty acids were the known natural ligands of PPAR- γ ,²¹ and few studies have demonstrated that omega-3 fatty acids decreased NF-kB activation by increasing PPAR- γ gene expression in a chain length-dependent manner.^{12,13} Being a member of the immunoglobulin superfamily with a high affinity for AGEs,²⁰ RAGE

Table 3. Advanced glycation end products and inflammatory cytokines at study ba omega-3 fatty acid supplements or placebo	okines at study baseline and after 3-month intervention in patients with diabetic nephropathy that received either
Placebo Group (n = 30)	Omega-3 fatty acids group (n=30) P

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	Baseline	End of Study	Change	r	Baseline	End of Study	Change	à	lime	Group	lime x Group
TNF-α, ng/L	19.0 ± 2.5	20.3 ± 5.8	1.3 ± 8.6	.42	19.1 ± 4.1	20.5 ± 4.5	1.4 ± 4.5	£.	.14	88.	96.
TNF-a receptor, ng/mL	21.0 ± 4.1	21.9 ± 3.4	0.9 ± 3.2	.13	19.7 ± 3.4	20.8 ± 4.1	1.1 ± 4.7	.24	.07	.14	.89
GDF15, ng/L	73.9 ± 8.8	73.6 ± 6.6	-0.3 ± 13.5	89.	77.1 ± 7.7	75.2 ± 9.3	-1.8 ± 10.9	.35	.49	.08	.63
AGEs, AU	25.8 ± 3.3	26.0 ± 2.0	0.2 ± 2.5	.67	27.0 ± 3.6	24.7 ± 2.7	-2.3 ± 2.8	< .001	.004	06.	.001
RAGEs, ng/mL	2.0 ± 0.2	2.1 ± 0.2	0.1 ± 0.3	.53	2.0 ± 0.4	1.9 ± 0.3	-0.1 ± 0.3	.02	.33	.03	.05
*TNE - indicator trans normalis factor - African definition and stadiusts. OE16 - annut differentiation factor 16, and BAC6 - annuts definition	rocio footor AC		otion and produc		E arouth difford	ntiation factor 16. 0		otor oduopo	and all continue	outors bao	

advanced glycation end products. 15; and RAGEs, receptor 15, growth differentiation factor בה end products; advanced glycation AGES. lactor-o.; -o. Indicates tumor necrosis Ż

Table 4. Adjusted Changes in Metabolic Variables in PatientsWith Diabetic Nephropathy Who Received Omega-3 Fatty AcidSupplements or Placebo*

Variables	Placebo Group (n = 30)	Omega-3 Group (n = 30)	Р
TNF-α, ng/L			
Model 1	1.2 ± 0.9	1.4 ± 0.9	.90
Model 2	1.2 ± 0.9	1.4 ± 0.9	.91
TNF-α receptor, ng/mL			
Model 1	1.3 ± 0.6	0.7 ± 0.6	.51
Model 2	1.3 ± 0.6	0.7 ± 0.6	.52
GDF15, ng/L			
Model 1	-2.1 ± 1.5	-0.1 ± 1.5	.30
Model 2	-2.1 ± 1.5	-0.1 ± 1.5	.28
AGEs, AU			
Model 1	-0.1 ± 0.3	-2.0 ± 0.3	< .001
Model 2	-0.1 ± 0.3	-2.0 ± 0.3	< .001
RAGEs, ng/mL			
Model 1	0.1 ± 0.1	-0.1 ± 0.1	.002
Model 2	0.1 ± 0.1	-0.1 ± 0.1	.003

*Model 1 is adjusted for baseline values of biochemical parameters, and model 2, for baseline values of biochemical parameters, age and baseline body mass index. TNF- α indicates tumor necrosis factor- α ; AGEs, advanced glycation end products; GDF15, growth differentiation factor 15; and RAGEs, receptor advanced glycation end products.

is associated with various diseases including Alzheimer disease, cardiovascular disease, and diabetic vasculopathy.²²

Supporting with our findings, taking 15 mL/d of either flaxseed oil among healthy young adults for 6 weeks did not decrease inflammatory markers.²³ In another study, low- and high-dose plant and marine (n-3) fatty acids did not influence plasma inflammatory markers in adults with metabolic syndrome after 8 weeks.²⁴ In addition, no significant effect on inflammatory factors was observed following the consumption of omega-3-enriched supplements of flaxseed and fish oil among firefighters for 12 weeks.²⁵ Similar findings were seen after omega-3 fatty acids supplementation from flaxseed oil among healthy volunteers for 12 weeks and healthy abdominally obese adults for 8 weeks.^{26,27} However, some researchers could observe the beneficial effects of omega-3 fatty acids supplementation on inflammatory cytokines. For instance, Perunicic-Pekovic and colleagues²⁸ demonstrated a significant decrease in serum TNF-a and interleukin-6 levels of hemodialysis patients treated with 2.4 g/d of omega-3 fatty acids from fish oil for 2 months. Moreover, circulating levels of TNF- α and intterleukin-1 β were reduced after

omega-3 fatty acids supplementation from fish oil in obese but not in normal-weight women for 4 weeks.²⁹ The discrepancies between our findings and those of previous reports might be explained by the origin of omega-3 fatty acids, the dosage of omega-3 fatty acids supplements used, the intervention time, the study participants, the quality of the supplements, their purity and bioavailability, and the time and period of administration.

Our study had some limitations. The main limitation of our study was the lack of measurements of circulating levels of omega-3 fatty acids at study baseline and at the end of the study due to budget limitations. The sample size of our study was low. Future studies with cross-over design, higher dose of omega-3 fatty acid, and bigger sample size are needed to confirm the validity of our findings in DN patients. In addition, we agreed that to determine other inflammatory cytokines (especially interleukin-6) in patients with DN is valuable. Actually, due to limited funding for research projects in developing countries, we did not evaluate these markers. Therefore, measurement of other inflammatory cytokines, biomarkers of oxidative stress, and HbA1c after intervention are warranted in future studies.

CONCLUSIONS

Overall, the current study demonstrated that omega-3 fatty acid supplementation for 12 weeks had favorable effects on serum AGEs and RAGEs levels among DN patients, but did not affect inflammatory cytokines.

CONFLICT OF INTEREST

None declared.

FINANCIAL SUPPORT

The study was supported by a grant (No 5979) from Kashan University of Medical Sciences.

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Received January 2016 Revised March 2016 Accepted April 2016